

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
24 January 2002 (24.01.2002)

PCT

(10) International Publication Number
WO 02/06452 A2

(51) International Patent Classification⁷: C12N 9/00

(74) Agent: SWABEY OGILVY RENAULT; Suite 1600,
1981 McGill College Avenue, Montreal, Québec H3A 2Y3
(CA).

(21) International Application Number: PCT/CA01/01032

(22) International Filing Date: 13 July 2001 (13.07.2001)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
60/218,842 18 July 2000 (18.07.2000) US

(71) Applicant (*for all designated States except US*): NATIONAL RESEARCH COUNCIL OF CANADA [CA/CA]; Corporate Services, Intellectual Property Services, 1200 Montreal Road, Building M-58, Room EG-12, Ottawa, Ontario K1A 0R6 (CA).

(81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

(72) Inventors; and

(75) Inventors/Applicants (*for US only*): IWAKI, Hiroaki [JP/CA]; 1625 West De Maisonneuve Boulevard, Suite 1508, Montreal, Québec H3H 2N4 (CA). HASEGAWA, Yoshie [JP/JP]; 8-45 Gojo-cho, Higashi-Osaka, Higashi-Osaka, Osaka 579-8042 (JP). LAU, Peter, C., K. [CA/CA]; 30 Piper's Crescent, Kirkland, Québec H9H 3J4 (CA).

Published:

— without international search report and to be republished upon receipt of that report

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: CLONING, SEQUENCING AND EXPRESSION OF A COMAMONAS CYCLOPENTANONE 1,2-MONOOXYGENASE-ENCODING GENE IN *ESCHERICHIA COLI*

(57) Abstract: Cyclopentanone 1,2-monooxygenase (CPMO) from *Comamonas* (previously *Pseudomonas*) sp. strain NCIMB 9872 carries out the second step of a degradation pathway that allows the bacterium to use cyclopentanol as a sole carbon source for growth. In the present invention there is reported the localization of the CPMO-encoding gene (*cpnB*) on a 4.3-kb *SphI* fragment, the determination of its sequence. The 550-amino acid CPMO polypeptide (M_r , 62,111) encoded by the gene was found to have 36.5% identity with the sequence of cyclohexanone 1,2-monooxygenase (CHMO) of *Acinetobacter* sp. strain NCIMB 9871. The 62-kDa CPMO was expressed in *E. coli* as an IPTG-inducible protein.

BEST AVAILABLE COPY

CLONING, SEQUENCING AND EXPRESSION OF A COMAMONAS
CYCLOPENTANONE 1,2-MONOOXYGENASE-ENCODING GENE IN
ESCHERICHIA COLI

TECHNICAL FIELD

5 The present invention relates to an isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, and expression vector and a transformed cell containing the isolates DNA.

BACKGROUND OF THE INVENTION

10 *Comamonas* (previously *Pseudomonas*) sp. NCIMB 9872 was one of the few microorganisms that have been characterized to produce a Baeyer-Villiger monooxygenase (BVMO; Griffin, M., et al., *Biochem. J.* **129**:595-603, 1972; Griffin, M., et al., *Eur. J. Biochem.* **63**:199-209, 1976; and Willetts, A., *Trends in Biotech.* **15**:55-62, 1997; for a recent review). BVMOs are flavoproteins that
15 mimic the classical Baeyer-Villiger organic chemical reaction which is a peracid-catalyzed oxidation of a ketone to an ester or lactone. The use of enzyme substitutes for the production of lactones in high yield and optical purity is an attractive feature in current trends of research and development toward replacing chemical methods with biological alternatives (Stinson, S.C., *Chem.*
20 *Eng. News*, 83-104, 1998). To date, the best characterized BVMO enzyme is that of cyclohexanone monooxygenase (CHMO) produced by *Acinetobacter* sp. NCIMB 9871 (Stewart, J.D., *Curr. Org. Chem.* **2**:195-216, 1998; Willetts, A., *Trends in Biotech.* **15**:55-62, 1997). This is also the only BVMO whose gene has been cloned and sequenced (Chen, et al., *J. Bacteriol.* **170**:781-789, 1988).
25 Recently, this valuable resource was used to engineer a "designer yeast" in a whole-cell approach to effect a variety of asymmetric Baeyer-Villiger oxidations (Stewart, J.D., et al., *J. Am. Chem. Soc.* **120**:3541-3548, 1998).

30 It would be highly desirable to be provided with a new CPMO having an increased enzymatic activity for growing cells in a medium containing cyclopentanol or cyclopentanone as sole carbon source.

SUMMARY OF THE INVENTION

One aim of the present invention is to provide a new CPMO having an increased enzymatic activity for growing cells in a medium containing cyclopentanol or cyclopentanone as sole carbon source.

5 In accordance with the present invention there is provided an isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, the isolated DNA being characterized by the ability to hybridize specifically with the complement of the DNA represented in SEQ ID NO:8 under stringent hybridization conditions.

10 Also in accordance with the present invention, there is provided an isolated DNA, wherein it codes for a cyclopentanone monooxygenase (CPMO), and contains:

- (1) the nucleic acid sequence of SEQ ID NO:8;
- (2) a sequence corresponding to said nucleic acid sequence in
15 the scope of the degeneration of the genetic code; or
- (3) a sequence hybridizing under stringent conditions with the sequence from (1) or (2), and still coding for cyclopentanone monooxygenase (CPMO).

20 Still in accordance with the present invention, there is provided an isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, said isolated DNA having SEQ ID NO:8.

The present invention further provides an isolated DNA expression vector encoding an enzymatically active cyclopentanone monooxygenase (CPMO)
25 comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.

In accordance with the present invention, there is also provided a recombinant vector comprising the isolated DNA as described above, wherein the isolated DNA encodes cyclopentanone monooxygenase.

In a preferred embodiment of the present invention, the isolated DNA has a nucleic acid sequence of SEQ ID NO:8 or which, due to the degeneracy of the genetic code, is a functional equivalent thereof.

Also in accordance with the present invention, there is provided a
5 recombinant vector containing one or more copies of a recombinant DNA described above.

The recombinant vector may be a prokaryotic vector. The recombinant vector may also be a plasmid.

Therefore, in accordance with the present invention, there is also
10 provided a biologically functional plasmid or viral DNA vector, which contains a DNA as described above.

The present invention also provide a host cell comprising a recombinant vector as described above.

Accordingly, there is also provided a cell transformed with a heterologous
15 DNA expression construct encoding an enzymatically active cyclopentanone monooxygenase (CPMO) comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.

The cell may be a prokaryotic cell or it may be *E. coli*.

20 Still in accordance with the present invention, there is also provided a purified cyclopentanone monooxygenase (CPMO) having :

- a) an amino acid sequence as set forth in SEQ ID NO:5;
- b) an amino acid sequence encoded by a nucleic acid sequence as set forth in SEQ ID NO:8; or
- 25 c) an amino acid sequence encoded by a nucleic acid sequence hybridizing to a nucleic acid sequence complementary to the nucleic acid sequence of step b) above under stringent conditions, said amino acid sequence encoded in step c) having a same activity as the amino acid sequence in a).

The present invention also provides a recombinant cyclopentanone monooxygenase (CPMO) having an enzymatic activity superior to the one from a native *Pseudomonas*, and more preferably twice superior.

5 The recombinant cyclopentanone monooxygenase (CPMO) may be prepared from *Comamonas* sp. NCIMB 9872. The recombinant cyclopentanone monooxygenase (CPMO) has preferably a sequence as set forth in SEQ ID NO:5.

10 A method for growing cells *in vitro* in presence of cyclopentanol or cyclopentanone as sole source of carbon, said method comprising the steps of:

- a) transforming a cell with the expression construct described above; and
- b) growing the cell of step a) under suitable conditions in a medium containing cyclopentanol or cyclopentanone as a sole
15 source of carbon.

To increase this gene potential, according to the invention it is reported herein the cloning of a cyclopentanone monooxygenase (CPMO)-encoding gene (*cpnB*) from *Comamonas* (*Pseudomonas*) sp. NCIMB 9872, the determination of its DNA and surrounding sequence and expression of CPMO
20 activity and protein in *E. coli*.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 illustrates the first two steps of cyclopentanol degradation by *Pseudomonas* sp. NCIMB 9872;

25 Fig. 2 illustrates the genetic organization in *Comamonas* sp. NCIMB 9872 in the *SphI* fragment containing cyclopentanone monooxygenase-encoding gene (*cpnB*) and additional open reading frames;

Fig. 3 illustrates an alignment of the amino acid sequence of the CPMO of *Comamonas* sp. NCIMB 9872 with that of CHMO from *Acinetobacter* sp.

NCIMB 9871 and a steroid monooxygenase (STMO) from *Rhodococcus rhodochrous*;

Fig. 4 illustrates a SDS-PAGE of crude extracts from *E. coli* (pCMP201); and

5 Fig. 5 illustrates CPMO-encoding gene, designated cpnB.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

Cloning of the *Comamonas* sp. NCIMB 9872 CPMO-encoding gene

Pseudomonas sp. NCIMB 9872 (henceforth strain 9872) identified as a *Comamonas* by 16S rDNA sequencing in this study, was purchased from the
10 National Collections of Industrial and Marine Bacteria Ltd (NCIMB, Aberdeen, Scotland) and grown at 30°C in Luria-Bertani (LB) broth (Sambrook, J., et al., Molecular cloning: a laboratory manual, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N. Y, 1989), or mineral salt medium (MSM), pH 7.0, containing 2 ml of cyclopentanone. The MSM recipe contains per liter: 1.0 g of
15 NH₄NO₃, 1.5 g of KH₂PO₄, 1.5 g of Na₂HPO₄, 0.2 g MgSO₄·7H₂O, 0.01 g of CaCl₂·2H₂O, 0.005 of FeSO₄·7H₂O, 0.002 g of MnSO₄·4H₂O and 0.1 g of yeast extract. Agar was added to 1.5% for plates. Genomic DNA of strain 9872 was prepared by the Marmur method (Marmur, J., *J. Mol. Biol.* 3: 208-218, 1961). At first, a Southern hybridization of DNA digested with *Bam*HI was carried out
20 using the *Acinetobacter* NCIMB 9871 CHMO-containing gene as probe. Since there was no positive result (hybridization conditions carried out at 65°C) the CPMO protein was purified in order to obtain an N-terminal amino acid sequence. The purification of CPMO protein from cyclopentanone-grown cells was according to Griffin and Turgill (Griffin, M., et al., *Eur. J. Biochem.* 63:199-
25 209, 1976). Using an automated protein sequencer (Perkin-Elmer model 477) a 40-residue amino-terminal sequence of the purified CPMO was obtained (Fig. 2). This sequence, longer by 11 amino acids, is in perfect agreement with that reported previously from the same organism (Willets, A., *Trends in Biotech.* 15:55-62, 1997). Two degenerate oligodeoxynucleotide primers (5'-
30 ACIACIATGA CIACNATGAC-3' (SEQ ID NO:1) and 5'-ARRTGRTAIA

RYTGRTA-3' (SEQ ID NO:2), corresponding to amino acids 2-8 and 35-40, respectively) were synthesized to amplify a 116-bp product from total DNA prepared from strain 9872. The PCR amplification was performed in a Perkin Elmer-Model 2400 Thermal Cycler™ and the amplification conditions were 94°C for 1 min, 50°C for 1 min and 72°C for 1 min for 30 cycles. The amplified product was cloned directly in the pXcmkn12 vector (Cha, J., et al., *Gene* **136**, 369-370, 1993), transformed in *E. coli* JM109 and the resulting plasmid was designated pCMP10. Before using the amplified product as a gene probe its nucleotide sequence was confirmed. Nucleotide sequencing was determined by the Taq DyeDeoxy terminator cycle sequencing kit and the ABI Prism 310 Genetic Analyzer (Perkin Elmer). Plasmid isolation was performed by the method of Birnboim and Doly (Birnboim, H. C., and J. Doly, *DNA. Nucleic Acids Res.* **7**:1513-1523, 1979).

In Fig. 2, Orf1 is most likely a transcriptional activator of the NtrC-type (Morett, E., L. Segovia, *J. Bacteriol.* **175**:6067-6074, 1993). The amino acid sequence of ORF1 (C-terminal 391-amino acids) showed 38-40% identity to equivalent regions of proteins such as NTRC_ECOLI (Nitrogen regulation protein NR(I) from *E. coli*; Miranda, et al., *The complete nucleotide sequence of the glnALG operon of Escherichia coli K12* **15**:2757-2770, 1987), ACOR_ALCEU (Acetoin catabolism regulatory protein from *Ralstonia eutropha*; Kruger, N., et al., *J. Bacteriol.* **179**:4391-4400, 1992). The amino acid sequence of ORF2, showing similarity to enzymes of the short-chain alcohol dehydrogenase family (Jornvall, H., et al., *Biochemistry* **34**: 6003-6013, 1995), is most homologous (45-46% identity) to a putative oxidoreductase CY39.16C of *Mycobacterium tuberculosis* (Swiss Prot sp:Q10855) and fadG3 of *M. tuberculosis* (GenBank accession number Z74025). For *Pseudomonas* sp. strain HI-201 the *lacZ*-Km^r cassette from pKOK6.1 (Kokotek, W., et al., *Gene* **84**: 467-471, 1989) was inserted into *cpnB* at the *Nsi*I site.

In Fig. 2, the following terms are defined as follows: t_{fd}, transcriptional termination sequence of phage fd; Km^r, kanamycin resistance gene, *lacZ*, gene encoding b-galactosidase. Genes and markers are indicated with arrows.

To clone the CPMO-containing gene, the DNA insert from pCMP10 was amplified, labeled by the digoxigenin-11-UTP system according to manufacturer's instructions (Boehringer Mannheim GmbH) and used to probe a Southern hybridization of strain 9872 genomic DNA digested with various restriction enzymes (*Bam*HI, *Eco*RI, *Hind*III, *Kpn*I, *Nhe*I, *Pst*I, *Sal*I, *Sph*I and *Xba*I). As a result, a single hybridizing band of ca 4.3-kb *Sph*I fragment was obtained. Conditions of hybridization were as before. Subsequently, a purified 4.0- to 4.5-kb size fraction of *Sph*I-cut total DNA separated on a 0.8 % agarose gel was ligated to *E. coli* plasmid pUC18, which had been linearized and dephosphorylated. A clone containing the 4.3-kb insert was screened by colony hybridization using the PCR product as a probe; this recombinant plasmid was designated pCMP200.

DNA sequence of the CPMO-encoding gene (*cpnB*) and the flanking region

Nucleotide sequencing of the CPMO-encoding gene was initiated by using a primer designed from the sequence of the PCR product cloned in pCMP10 and further extended using oligonucleotides derived from the new sequence. Both DNA strands of the *Sph*I fragment were sequenced and found to consist of 4281 base pairs (bp). The sequence was analyzed by GENETYX-Mac (Software Development Co., Ltd. Chiba, Japan) and the BLAST program (Altschul, S. F., et al., *Nucleic Acids Res.* **25**:3389-3402, 1997). As a result three open reading frames (ORFs) arranged in the same direction were predicted (Fig. 2). The nucleotide sequence of the 1650-bp ORF encoding CPMO is preceded by a partial ORF1 (1173-bp) coding for the C-terminus of an NtrC-type transcriptional activator (Miranda, et al., *The complete nucleotide sequence of the glnALG operon of Escherichia coli K12* **15**:2757-2770, 1987) and by a complete ORF2 (750-bp) coding for a homolog of the short-chain dehydrogenases/reductases (Jornvall, H., et al., *Biochemistry* **34**: 6003-6013, 1995). The two intergenic regions are 244-bp and 32-bp, respectively. The CPMO-encoding gene is referred to *cpnB* (cyclopentanone and *B* designates the second step of the degradation pathway, see Fig. 1) hereafter. In Fig. 5, the CPMO-encoding gene starts at nucleotide position 1822 and ends 3471 that does not include the stop codon. Accordingly, the boundary of *cpnA* is 3507-

4256. The partial open reading frame preceding *cpnB* is from 1-1174.

Fig. 1 has been adapted from Griffin, M., et al. (Griffin, M., et al., *Biochem. J.* **129**:595-603, 1972). The designated genes are: *cpnA* encoding cyclopentanol dehydrogenase; *cpnB* encoding cyclopentanone 1,2-
5 monooxygenase (CPMO). An alternative name for 5-valerolactone is 5-pentanolide. Subsequent reaction steps are the formation of 5-hydroxyvalerate, 5-oxovalerate, glutarate and finally acetyl CoA.

The amino acid sequence of the CPMO enzyme consists of 550 residues (Fig. 3). This sequence shows 36.5% identity and an additional 13.6% amino
10 acid similarity with the 543-residue CHMO of *Acinetobacter* sp. strain NCIMB 9871. An equally related protein (549 amino acids; 37.3% identity and 12.4% similarity) is the putative steroid monooxygenase (STMO) of *Rhodococcus rhodochrous* (Morii, S., et al., GenBank accession number AB010439, 1998). The latter enzyme carries out the oxidation of progesterone to produce
15 testosterone acetate. A CLUSTAL alignment of these three sequences gave 24.6% positional identity (Fig. 3).

In Fig. 3, asterisks indicate identical amino acids, dots indicated similar amino acids and dashes indicate gaps introduced to maximize the alignment. The amino-terminal peptide sequence confirmed by Edman degradation is
20 underlined. The locations of the consensus FAD fingerprint sequences as described by Eppink et al. (Eppink, et al., *Prot. Sci.* **6**:2454-2458, 1997) are as indicated. The conserved GD motif found in flavoprotein hydroxylases as a second FAD fingerprint is also indicated. Not shown is the DG motif of flavoprotein hydroxylases which has the sequence of chhhssDGxcSxhR. Lower
25 case letters identify certain residues types: h, hydrophobic residues, s, small residues, c, charged residues, and x, any residues. Note that a DG doublet is present in CPMO and STMO sequence.

A notable sequence motif present in CPMO and related proteins is the FAD-binding fingerprint (GXGXG) that is similar to those found in flavoprotein
30 hydroxylases (Eppink, et al., *Prot. Sci.* **6**:2454-2458, 1997). Flavoprotein hydroxylases (eg. phenol hydroxylase, the structure is now known; Enroth, C.,

et al., *Structure* 6:605-617, 1998) are monooxygenases that catalyze the insertion of one atom of molecular oxygen into the substrate using NAD(P)H as electron donor. These proteins possess a conserved "Asp-Gly (DG)" motif for both FAD and NAD(P)H binding in between two fingerprint motifs for the FAD binding (fingerprint 1: GXGXXG; fingerprint 2: Gly-Asp [GD] motif). Sequence motifs in CPMO, STMO and CHMO differ from those in flavoprotein hydroxylases by having a repeated GXGXXG motif (amino acids 24 to 33 and 193 to 202 in CPMO numbering). The possibility that the second FAD fingerprint in CPMO and related proteins fulfils a dual role of FAD and NADPH binding awaits structural determination of a representative member of this family of proteins. It is reasonable to assume that a different mechanism in catalysis is reflected in the motifs seen in the two classes of proteins.

Expression of *cpnB* gene in *E. coli*

Two primers of the following sequence were synthesized to amplify the *cpnB* gene and the resultant 1.7-kb DNA fragment was cloned in the pSD80 plasmid to yield pCMP201. Plasmid pSD80 is a third generation derivative of the commercially available pKK223-3 vector (Pharmacia) that contains a *tac* promoter upstream of the multiple cloning site (MCS), an *unc* terminator sequence downstream of the MCS, and *lacI^q* elsewhere on the plasmid (Smith, S.P., et al., *Biochemistry* 35:8805-8814, 1996). The primers were: 5'-AAAAGGCCTG AACTTCAATT ATTAGGAGA C-3' (SEQ ID NO:3) and 5'-AAAACTGCAG GAGTTGCACA ACAGAGTCTT AG-3' with built-in *StuI* and *PstI* restriction sites (underlined), respectively, to facilitate cloning at the compatible sites (*SmaI* and *PstI*) of the pSD80 vector. Vent DNA polymerase (New England BioLabs, Beverly, MA) was used and the amplification conditions were 94°C for 1 min, 50°C for 1 min, and 72°C for 1 min, for 30 cycles. The amplified DNA fragment was purified from an agarose gel and digested with *StuI* and *PstI*. One of the resulting recombinant plasmids was designated pCMP201. By DNA sequencing it was established that no mutation had been introduced in the *cpnB* gene during PCR amplification.

Fig. 4 shows the production of a 60-kDa protein in a Coomassie blue-stained SDS-polyacrylamide gel of the crude protein extract prepared from *E. coli* JM109 (pCMP201) cells that were induced by 0.1 mM isopropyl-beta-D-thiogalactopyranoside (IPTG). The cells were induced at an absorbance (A₆₀₀) of 0.4 to 0.5 and the induction period was up to 4 hr. The observed molecular mass was in agreement with the predicted size of the 62-kDa CPMO. In the absence of IPTG, this protein band was not produced. Also, the CPMO enzyme activity was observed only in those cells grown in the presence of IPTG. CPMO activity was assayed at 25°C by measuring a decrease in absorbance at 340 nm in 50 mM phosphate buffer (pH 7.8) containing 1 µmol of cyclopentanone, 0.2 µmol of NADPH, and the crude enzyme extract prepared from *E. coli* JM109 (pCMP201). These cells were cultivated in 100 ml of LB medium containing 100 µg/ml of ampicillin at 25°C. The IPTG-induced cells were harvested by centrifugation, washed in 50 mM phosphate buffer (pH 7.2), resuspended in 1/20 volume of same buffer, and sonicated by four-20 sec bursts with a Braun-Sonifier™ 250 apparatus. After centrifugation for 30 min at 18,000 x g and at 4°C, the supernatant was used for determination of enzyme activity. One unit (U) of activity is defined as the amount of enzyme required to convert 1 µmol of substrate in 1 min. Protein concentration was determined by the method of Bradford (Bradford, M. M., *Anal. Biochem.* **72**: 248-254, 1976). As a result the specific activity of the CPMO enzyme was found to be 0.28 U/mg. The specific activity of CPMO in the native *Pseudomonas* was reported to be 0.11 U/mg (Griffin, M., et al., *Biochem. J.* **129**:595-603, 1972).

In Fig. 4, lane 1 has been loaded with extracts of IPTG-induced *E. coli* and lane 2 has been loaded with extracts of *E. coli* in absence of IPTG. M means molecular weight markers as indicated in kilo daltons. The arrow indicates the production of the desired 60-kDa protein.

Inactivation of *cpnB* gene

Pseudomonas sp. strain HI-201 was constructed by chromosomal inactivation of the *cpnB* gene using a *lacZ*-Km^r cassette from the mobilizable pKOK6.1 vector (Kokotek, W., et al., *Gene* **84**: 467-471, 1989). In pKOK6.1 the

lacZ gene is promoterless and in addition to Km^r it is ampicillin resistant (Ap^r). The *lacZ*-Km^r cassette was excised as a *Pst*I-fragment and inserted into the *Nsi*I site within the *cpnB* gene in pCMP200, yielding pCMP202. Electroporation of this plasmid into 9872 cells was carried out in the Gene Pulser™ (BioRads) and the parameters of electroporation were 2.5 kV, 25 uF and 200 ohm. The cells were initially washed with 1mM HEPES buffer and resuspended in 1mM HEPES containing 10% glycerol. Km^r colonies were selected on LB plates containing Km (250 µg/ml). To select for double crossover mutants, a second screening on LB plates containing Ap (300 µg /ml) was carried out. The inactivation of *cpnB* (Fig. 2), was confirmed by PCR. The resulting mutant HI-201 was found not to be able to grow on cyclopentanol or cyclopentanone as a sole carbon and energy source. This result indicated that *cpnB* is essential for the degradation of cyclopentanol and it appeared that there was only one copy of the *cpnB* gene in strain 9872.

As expected of a flavoprotein the amino acid sequence of CPMO contains motifs of FAD fingerprints similar to those found in flavoprotein hydroxylases.

Nucleotide sequence accession number

The DNA sequence of the 4,281-bp *Sph*I fragment has been submitted to DDBJ and assigned accession number AB022102. The release of this data awaits the inventors' authorization.

While the invention has been described in connection with specific embodiments thereof, it will be understood that it is capable of further modifications and this application is intended to cover any variations, uses, or adaptations of the invention following, in general, the principles of the invention and including such departures from the present disclosure as come within known or customary practice within the art to which the invention pertains and as may be applied to the essential features hereinbefore set forth, and as follows in the scope of the appended claims.

WHAT IS CLAIMED IS:

1. An isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, the isolated DNA being characterized by the ability to hybridize specifically with the complement of the DNA represented in SEQ ID NO:8 under stringent hybridization conditions.
2. An isolated DNA, wherein it codes for a cyclopentanone monooxygenase (CPMO), and contains:
 - (a) the nucleic acid sequence of SEQ ID NO:8;
 - (b) a sequence corresponding to said nucleic acid sequence in the scope of the degeneration of the genetic code; or
 - (c) a sequence hybridizing under stringent conditions with the sequence from (1) or (2), and still coding for cyclopentanone monooxygenase (CPMO).
3. An isolated DNA encoding a cyclopentanone monooxygenase (CPMO), or an enzymatically active portion thereof, said isolated DNA having SEQ ID NO:8.
4. An isolated DNA expression vector encoding an enzymatically active cyclopentanone monooxygenase (CPMO) comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.
5. A recombinant vector comprising the isolated DNA of any one of claims 1 to 3, wherein the isolated DNA encodes cyclopentanone monooxygenase.
6. The recombinant vector of claim 5, wherein the isolated DNA has a nucleic acid sequence of SEQ ID NO:8 or which, due to the degeneracy of the genetic code, is a functional equivalent thereof.
7. A recombinant vector, wherein it contains one or more copies of a recombinant DNA according to claim 1, 2 or 3.
8. A recombinant vector according to claim 4, 5, 6 or 7, wherein it is a prokaryotic vector.

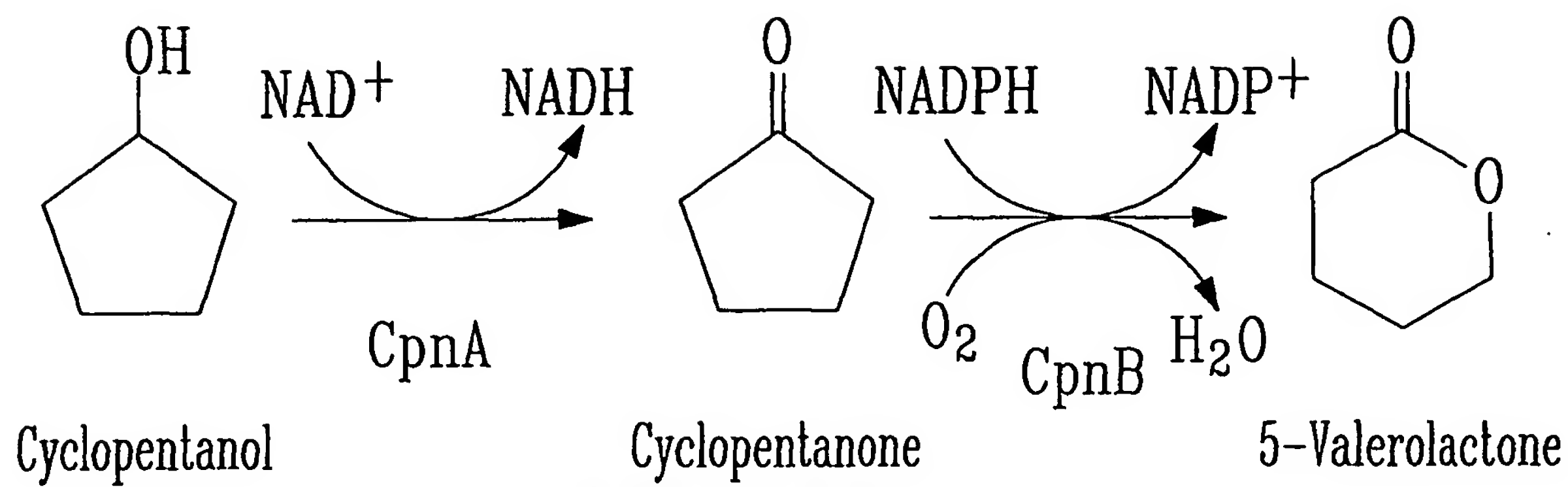
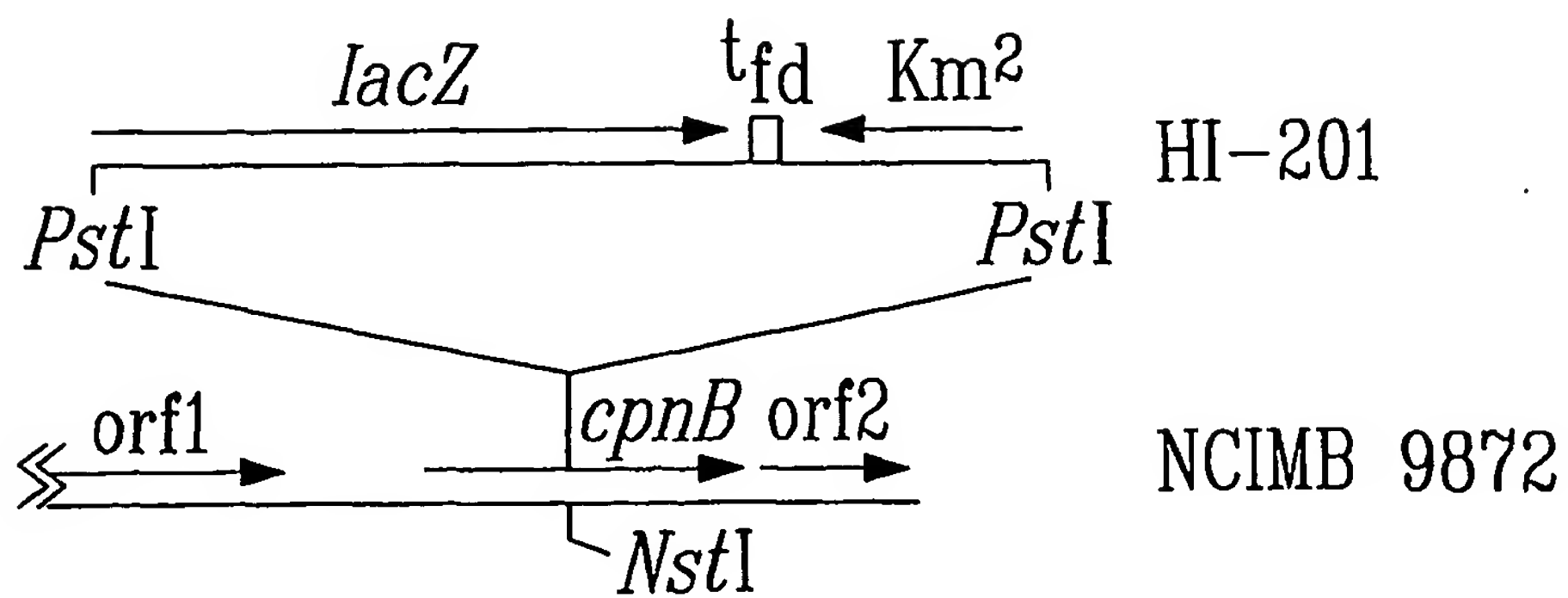
9. A recombinant vector according to claim 4, 5, 6 or 7, wherein it is a plasmid.
10. Biologically functional plasmid or viral DNA vector, which contains a DNA as defined in claim 1, 2 or 3.
11. A host cell comprising a recombinant vector of any one of claims 4 to 9.
12. A cell transformed with a heterologous DNA expression construct encoding an enzymatically active cyclopentanone monooxygenase (CPMO) comprising a DNA characterized by a sequence as set forth in SEQ ID NO:8, or a portion thereof, said portion encoding said CPMO, in expressible form.
13. A cell as defined in claim 12, which is a prokaryotic cell.
14. A cell as defined in claim 12, which is *E. coli*.
15. A purified cyclopentanone monooxygenase (CPMO) having :
 - (a) an amino acid sequence as set forth in SEQ ID NO:5;
 - (b) an amino acid sequence encoded by a nucleic acid sequence as set forth in SEQ ID NO:8; or
 - (c) an amino acid sequence encoded by a nucleic acid sequence hybridizing to a nucleic acid sequence complementary to the nucleic acid sequence of step b) above under stringent conditions, said amino acid sequence encoded in step c) having a same activity as the amino acid sequence in a).
16. A recombinant cyclopentanone monooxygenase (CPMO) having an enzymatic activity superior to the one naturally occurring.
17. The recombinant cyclopentanone monooxygenase (CPMO) of claim 16, wherein said CPMO is prepared from *Comamonas* sp. NCIMB 9872.
18. The recombinant cyclopentanone monooxygenase (CPMO) of claim 16, wherein said CPMO has a sequence as set forth in SEQ ID NO:5.

19. The recombinant cyclopentanone monooxygenase (CPMO) of claim 16, wherein said CPMO has an enzymatic activity twice superior to that of a CPMO from a native *Pseudomonas*.

20. A method for growing cells in vitro in presence of cyclopentanol or cyclopentanone as sole source of carbon, said method comprising the steps of:

- (a) transforming a cell with the expression construct of claim 4, 5, 6, 7, 8 or 9; and
- (b) growing the cell of step a) under suitable conditions in a medium containing cyclopentanol or cyclopentanone as a sole source of carbon.

1/10

FIG. 1FIG. 2

2/10

GxGxxG

CHMO 1:-----MSQKMDFDAIVIGGGFGGLYAVKKLRDELELKVQAFDKATDVAGT 45

STMO 1:MNGQHPRSVVTAPDATTGTTSDV VVGAGIAGLYAIHRFRSQ-GLTVRAFEAASGVGGV 59

CPMO 1:MTTMTTTEQLGMNNSVNDKLDVLLIGAGFTGLYQLYHLRKL-GYKVHLVDAGADIGGI 59

. * . . . * * . *** . . . * * . . . *

CHMO 46:WYWNRYPGALTDTEHLYCYSWDKELLQSLEIKKKYVQGPVVRKYLOQVAEKHDLKKSQ 105

STMO 60:WYWNRYPGARCDVESIDYSYSFSPELEQEWNWSEKYATQPEILAYLEHVADRFDLRRDIR 119

CPMO 60:WHWNCYPGARVDTHCQIYQYSI-PELWQEFNWKEFPNWAQMREYFHFADKKLDLSKDIS 118

* . ** ***** * * ** * * * . . . * . . ** .

CHMO 106:FNTAVQSAHYNEADALWEVTTEYGDKYTARFLITALGLLSAPNLPNIKGINQFKGELHHT 165

STMO 120:FDTRVTSAVLDEEGLRWTVRTDRGDEV SARFLVVAAGPLSNANTPAFDGLDRFTGDIVHT 179

CPMO 119:FNTRVQSAVFDEGTREWTVRSIGHQPIQARFVIANLGFGASPSTPNVDGIETFKGQWYHT 178

* . * * * * . * * * . . *** . . * . * * . * * . *

GxGxxG

CHMO 166:SRWPDD-VSFEGKRVGVIGTGSTGVQVITAVAPLAKHLTVFQRSAQYSVPIGNDPLSEED 224

STMO 180:ARWPHDGVDFGTGKRVGVIGTGSSGIQSIPIIAEQAEQLFVFQRSANY SIPAGNVPLDDAT 239

CPMO 179:ALWPQEGVNMAGKRVAIIGTGSSGVQVAQEAALDAKQVTVYQRTPNLALPMHQKQLSAED 238

. ** * . ***** . ***** . * * * * * * * . . . * . *

CHMO 225:VKKIKDNYDKSLGWCMNSALAFALNESTVPAMSVSAEERKAVFEKAWQTGGGFRFMFETF 284

STMO 240:RAEQKANYAERRRLSRESGGGSPHRPHPKSALEVSEEERRAVYEERWKLGG--VLFSKAF 297

CPMO 239:NLRMKPELPAAFERRGKCFAGFDFDFIAKNATELSAAERTEILEELWNAGG-FRYWLANF 297

* . . * . * * * . * . * * * . *

FILE - 3A

3/10

CHMO 285:GDIATNMEANIEAQNFIFKGKIAEIVKDPAIAQKLMPOD---LYAKRPLCDSGYNTFNR 340

STMO 298:PDQLTDPAANDTARAFWEEKIRAVVDDPAVAELLTPKDH--AIGAKRIVTDSGYETYNR 355

CPMO 298:QDYLFDDKANDYVVEFWRDKVRARIKDPKVAEKLAPMKKPHPYGAKRPSLEQWYEIFNQ 357

* . ** * * . ** . * * *** . ** . *

CHMO 341.DNVRLEDVKANPIVEITENGVKLENGDFVELDMLICATGFDAVDGNYVRMDIQKNGLAM 400

STMO 356:DNVELVDLRSTPIVGMDGTGIVT-TGAHYDLDMIVLATGFDAMTGS�DKLEIVGRGGRTL 414

CPMO 358:NNVTLDVNETPVLRITEKGIVT-AEGEAEFDLIVFATGFDAVTGGITSIDFRNNQGSF 416

. ** * * . * . . * * . . * . . * . . . * . .

CHMO 401:KDYWKEGPSSYMGVTVNNYPNMFVLGPNGP--FTNLPPSIESQVEWISDTIQYTVENNV 458

STMO 415:KETWAAGPRTYLGLGIDGFPNFFNLTGPGSPSVLANMVLHSELHVDWVADAIAYLDARGA 474

CPMO 417:KDVWSDGIRTQLGVATAGFPNLLFGYGPQSPAGFCNGPSSAEYQGDLLIQLMNYLRDNNI 476

* . * * . * . . ** . . ** * . * * *

CHMO 459:ESIEATKEAEEQWTQTCANIAEMTLFPAQSWIFGANIPGKKNTVYFYLGGLKEYRTCAS 518

STMO 475:AGIEGTPEAVADWVEECRNRAEASLLNSANSWYLGANIPGRPRVFMPFLGGFGVYREIIT 534

CPMO 477:SRIEAQSEAQEEWSKLIADFWDSSLFPRAKSWYQGSNIPGKKVESLNFPLGLPTYISKFN 536

** ** . * . . . * . * . ** * . **** . . * . *

CHMO 519:NCKNHAYEGFDIQLQRSDIKQPANA 543 SEQ ID NO:7

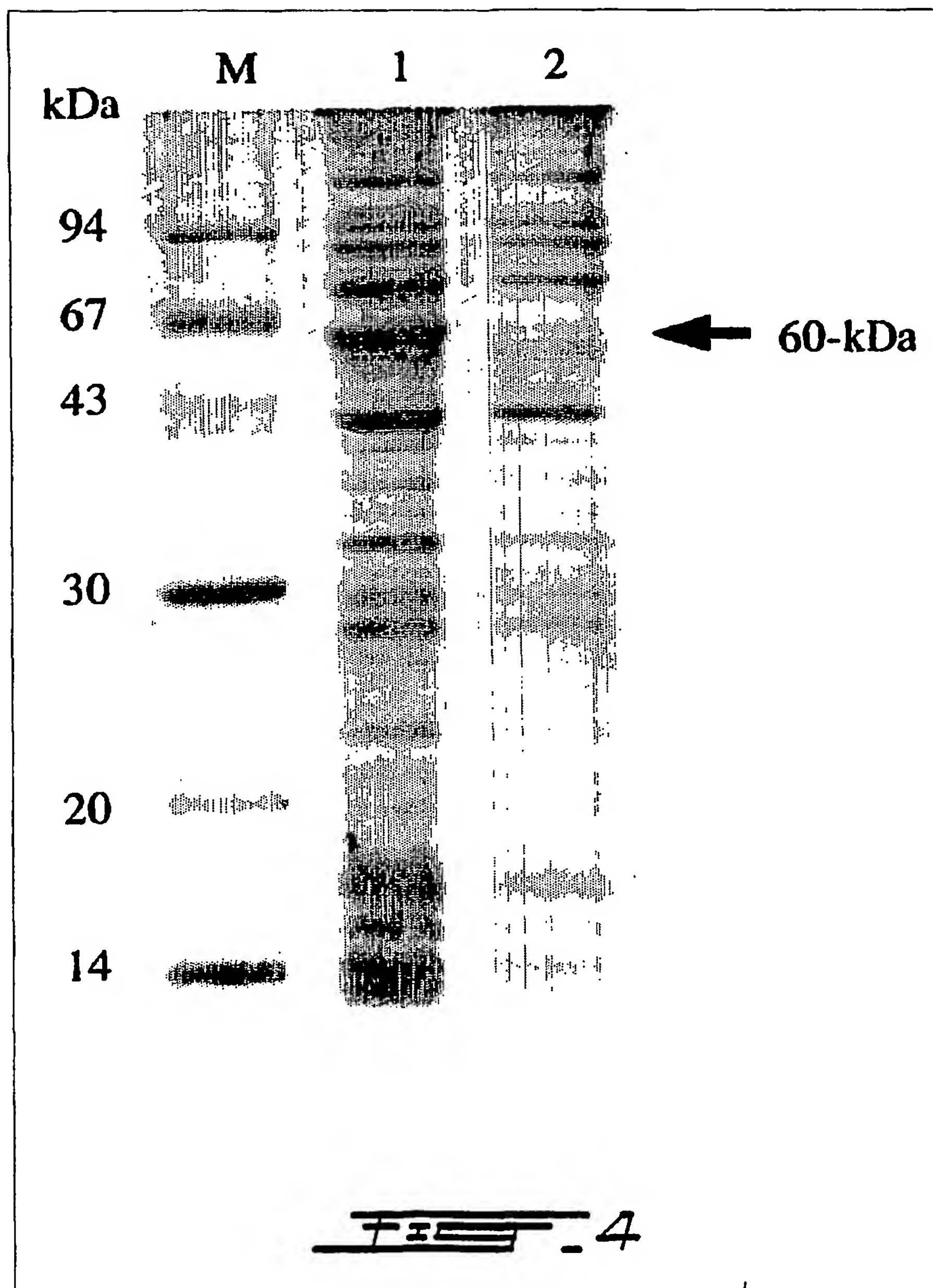
STMO 535:EVAESGYKGFAILEG----- 549 SEQ ID NO:6

CPMO 537:ESAEKGYAGFSLAS----- 550 SEQ ID NO:5

. . * ** .

FILED 38

4/10



5/10

	10	20	30	40	50	60
1	GCATGCGATAGCGGCACGTTGGTGGGCGGGCGAGCGCGGCCACGAGCGCGTCTCTCGCTC					
	CGTACGCTATCGCCGTGCAACCACCCGCCCGCTCGCGCCGGTGCTCGCGCAGAGAGCGAG					
61	CACCGACATCACATCGCCGGTCACCGGCGAGAACATTGGCAAGATCCTGCGCTTTTCGCC					
	GTGGCTGTAGTGTAGCGGCCAGTGGCCGCTCTTGTAACCGTTCTAGGACGCGAAAAGCGG					
121	GCTGAAGCCGCAGGGCTACGAGAGAATTTTTCAGTCCGGCCTGGCGGACTCGACCCAGCA					
	CGACTTCGGCGTCCCGATGCTCTCTTAAAAAGTCAGGCCGGACCGCCTGAGCTGGGTCTG					
181	CACCCCCAGGATGGTGCGTTTCGGACAGTTCCTGTCCTGTGATCCAGAGACCATCAACAC					
	GTGGGGGTCTTACCACGCAAAGCCTGTCAAGGACAGGACACTAGGTCTCTGGTAGTTGTG					
241	CCTGGAGACATTGGAGCGGATCGCGCGCGCCGACGTCAATGTCTTGCTGCATGGAGAGAC					
	GGACCTCTGTAACCTCGCCTAGCGCGCGCGGCTGCAGTTACAGAACGACGTACCTCTCTG					
301	GGGCACGGGCAAAGAGCTGATAGCTCGCCACATTCATGTCGCGAGCCGCCGGCGGGATGC					
	CCCGTGCCCGTTTCTCGACTATCGAGCGGTGTAAGTACAGCGCTCGGCGGCCGCCCTACG					
361	GCCCTACCTCGCCATCAACTGCGGGGCAATCAGCTCGGAGTTGCTGGAGAGTACTTTTTT					
	CGGGATGGAGCGGTAGTTGACGCCCCGTTAGTCGAGCCTCAACGACCTCTCATGAAAAAA					
421	TGGCTATGTGCGCGGAGCATTCTCCGGGGCAGATCCCAAGGGCCGCGCCGGCTACTTTGA					
	ACCGATACACGCGCCTCGTAAGAGGCCCGTCTAGGGTTCCCGGCGCGGCCGATGAAACT					
481	ATCGGTGGGCGAAGGCACCTTGTTTCTGGACGAGATCGGCGAGCTGCCCTTGGCCATGCA					
	TAGCCACCCGCTTCCGTGGAACAAAGACCTGCTCTAGCCGCTCGACGGGAACCGGTACGT					
541	GGCGGCATTGTTGCGCGTGCTGGAAGACGGAAGCTTTCTGCGTGTGGGCTCATCGACCCC					
	CCGCCGTAACAACGCGCACGACCTTCTGCCTTCGAAAGACGCACACCCGAGTAGCTGGGG					
601	GCAGCGCGCCTCGTGCCGCATTATCGCGGCCACGCACCGCAATCTGGAGGAACTCATCGC					
	CGTCGCGCGGAGCACGGCGTAATAGCGCCGGTGCGTGGCGTTAGACCTCCTTGAGTAGCG					
661	ACAGGGCCTGTTTCGCCAGGATCTCTACTACCGTCTCAAGATCGTTCAGAAAAGGCTCAA					
	TGTCCCGGACAAAGCGGTCCTAGAGATGATGGCAGAGTTCTAGCAAGTCTTTTCCGAGTT					

FIG. 5A


6/10

721 GCCGATACGGGAACGCACCTGTGACATCGCCTTGCTGGCAGAGCAATTCAAATCTGCACT
CGGCTATGCCCTTGCGTGGACACTGTAGCGGAACGACCGTCTCGTTAAGTTTAGACGTGA
781 GGCGCAAAGCACCAGATCCCGAACGTGCAGATCCACCCCGAGGCCATGGCAGTCATGGA
CCGCGTTTTTCGTGGTCTAGGGCTTGACGTCTAGGTGGGGCTCCGGTACCGTCAGTACCT
841 GCGCTACCAGTGGCCGGGCAATGCGCGCGAAATCCGCAATGTGATGGAAGCCGCCCTGAT
CGCGATGGTCACCGGCCCCGTTACGCGCGCTTTAGGCGTTACACTACCTTCGGCGGGACTA
901 CTGCTCCGATGGTGAAATCACGCTCGCCAGCCTGCCCCCGGAAGTATCTGAGAACTCCAC
GACGAGGCTACCACTTTAGTGCGAGCGGTTCGGACGGGGGCCTTCATAGACTCTTGAGGTG
961 CTATCCTCTGCAAAGCCGGGTGGCCGAAGCGAATAGCGAGATGCCTCCCGTCTCCGGCAA
GATAGGAGACGTTTCGGCCCACCGGCTTCGCTTATCGCTCTACGGAGGGCAGAGGCCGTT
1021 CGACTATGAACGGCAACTCATCGTGGGTTTGCTGCGCAAGTACCGAAAGGTCAATCATGT
GCTGATACTTGCCGTTGAGTAGCACCCAAACGACGCGTTCATGGCTTTCCAGTTAGTACA
1081 GGCAGGGGCCCTGGGCCTCGCGCGCTCCACGCTCTATCGGAAATTCGCAGACTTGGGCAT
CCGCTCCCGGGACCCGGAGCGCGCGAGGTGCGAGATAGCCTTTAAGCGTCTGAACCCGTA
1141 TGACCAACGCGAGTATGTCAATGATCTGTCCGATTAACACTGCTGCGCCCTGCGCTTGCG
ACTGGTTGCGCTCATAAGTTACTAGACAGGCTAATTGTGACGACGCGGGACGCGAACGC
1201 GCAACGCAGTATCGTCGGAACGCGGGCAACGCTAAAGGCAGCCGCCGTCGGCATCGGTTG
CGTTGCGTCATAGCAGCCTTGCGCCCGTTGCGATTTCCGTCGGCGGCAGCCGTAGCCAAC
1261 CCGCCCGGTGTTGTTTGTGCGCTGAGACCGCCTGCAGCACCAGCCCCTCCATTCCTCGTG
GGCGGGCCACAACAACAGCCGACTCTGGCGGACGTCGTGGTCGGGGAGGTAAGGAGCAC
1321 TCTAGATCTCCTTGCAACGCTGTCACAGCGTGCTTTTTTCGTGGCGCGGGACTTCTCGCG
AGATCTAGAGGAACGTTGCGACAGTGTCGCACGAAAAAAGCACCGCGCCCTGAAGAGCGC
1381 ATGAGGGCCTGCAAGGGCGGCGCATCAAACAAGCGCCCTGTCCTCAGGACGTTTGCGCC
TACTCCCGGACGTTCCCGCCGCGTAGTTTTGTTGCGGGACAGGAGTCCTGCAAACGCGG
1441 GTCGCTGGCTATGGGCAGTCTGCATGATCCGCTTGGGCTTGCCCACCATGCGCGCGACTG

Fi - 5B

7/10

CAGCGACCGATACCCGTCAGACGTACTAGGCGAACCCGAACGGGTGGTACGCGCGCTGAC
1501 CGCTCTTTCCTGTTGGGCGCATCGTCATCCCGACCTTCCTGAAATCCATTTCCACAGGGC
GCGAGAAAGGACAACCCGCGTAGCAGTAGGGCTGGAAGGACTTTAGGTAAAGGTGTCCCG
1561 CGGCCTGAGCCAGACAGCGATCGGCAGGCTCTCGCATCTCGGACAGTGGTGCGCTACACC
GCCGGACTCGGTCTGTGCTAGCCGTCCGAGAGCGTAGAGCCTGTCACCACGCGATGTGG
1621 ACAGGACACAAGTGTCCGGCGCGTGGCACGCAGCGGCCTTTGCTGCGCACGATGCGTGCT
TGTCCTGTGTTACAGGCCGCGCACCGTGCGTCGCCGGAACGACGCGTGCTACGCACGA
1681 TTTGGCACATGGAAACCCTCGGTTTTTCCGAACCAGAGACCTGGCACAAGTCTTGATAAC
AAACCGTGTACCTTTGGGAGCCAAAAAGGCTTGGTCTCTGGACCGTGTTCAGAACTATTG
1741 TCCTGAGTGACTGATGCAACGCATCTCAGTCCGGGAGCGTGGACGACACGCCCCCTGAACT
AGGACTCACTGACTACGTTGCGTAGAGTCAGGCCCTCGCACCTGCTGTGCGGGGACTTGA
1801 TCAATTATTTAGGAGACCCATATGACCACCATGACCACCATGACCACCGAACAACCTCGGC
AGTTAATAAATCCTCTGGGTATACTGGTGGTACTGGTGGTACTGGTGGCTTGTTGAGCCG
1861 ATGAACAACCTCTGTCAATGACAAGCTTGACGTTTTGCTCATCGGCGCCGGCTTCACCGGT
TACTTGTTGAGACAGTTACTGTTGGAAGTGCAAAACGAGTAGCCGCGGCCGAAGTGGCCA
1921 CTCTACCAGCTCTATCACCTGCGCAAGCTGGGCTACAAGGTTTCATCTCGTCGACGCCGGT
GAGATGGTCGAGATAGTGGACGCGTTCGACCCGATGTTCCAAGTAGAGCAGCTGCGGCCA
1981 GCCGATATTGGCGGGATCTGGCATTTGGAAGTGTACCCCGGAGCGCGTGTGGATACCCAC
CGGCTATAACCGCCCTAGACCGTAACCTTGACGATGGGGCCTCGCGCACACCTATGGGTG
2041 TGCCAGATCTACCAGTACTCCATTCCAGAGTTGTGGCAGGAGTTCAACTGGAAAGAGCTG
ACGGTCTAGATGGTCATGAGGTAAGGTCTCAACACCGTCCTCAAGTTGACCTTTCTCGAC
2101 TTCCCTAACTGGGCGCAAATGCGCGAGTATTTCCATTTTGCCGACAAGAAGCTCGACCTG
AAGGGATTGACCCGCGTTTACGCGCTCATAAAGGTAAAACGGCTGTTCTTCGAGCTGGAC
2161 AGCAAGGACATCAGCTTCAACACCCGTGTGCAGTCGGCCGTCTTTGACGAAGGCACACGC
TCGTTCCCTGTAGTCGAAGTTGTGGGCACACGTCAGCCGGCAGAACTGCTTCCGTGTGCG



8/10

2221 GAATGGACGGTACGCTCGATCGGACACCAGCCGATCCAGGCCAGGTTTCGTCATCGCCAAC
CTTACCTGCCATGCGAGCTAGCCTGTGGTCGGCTAGGTCCGGTCCAAGCAGTAGCGGTTG
2281 CTTGGCTTCGGTGCCAGCCCCAGCACGCCCAATGTCGATGGCATCGAGACATTCAAAGGC
GAACCGAAGCCACGGTCGGGGTCGTGCGGGTTACAGCTACCGTAGCTCTGTAAGTTTCCG
2341 CAGTGGTATCACACTGCTCTGTGGCCCCAGGAAGGCGTGAACATGGCCGGCAAGCGCGTG
GTCACCATAGTGTGACGAGACACCGGGGTCCTTCCGCACTTGTACCGGCCGTTTCGCGCAC
2401 GCCATCATTGGCACCGGCTCCAGCGGGGTCCAGGTCGCCCAGGAGGCTGCCCTTGATGCG
CGGTAGTAACCGTGGCCGAGGTGCGCCCAGGTCCAGCGGGTCCTCCGACGGGAACCTACGC
2461 AAACAGGTGACGGTGTACCAGCGCACCCCCAACCTGGCCTTGCCCATGCATCAGAAGCAG
TTTGTCCACTGCCACATGGTCGCGTGGGGGTGGACCGGAACGGGTACGTAGTCTTCGTC
2521 CTCAGCGCCGAGGACAATCTGCGCATGAAGCCCGAGCTTCCCGCAGCGTTCGAGAGACGC
GAGTCGCGGCTCCTGTTAGACGCGTACTTCGGGCTCGAAGGGCGTCGCAAGCTCTCTGCG
2581 GGCAAGTGCTTCGCCGGCTTCGACTTCGACTTCATCGCCAAGAACGCGACCGAGCTGTCC
CCGTTACGAAGCGGCCGAAGCTGAAGCTGAAGTAGCGGTTCTTGCGCTGGCTCGACAGG
2641 GCTGCGGAGCGCACAGAGATCTTGGAAGAGCTGTGGAACGCCGGCGGCTTCCGCTACTGG
CGACGCCTCGCGTGTCTCTAGAACCTTCTCGACACCTTGCGGCCGCCGAAGGCGATGACC
2701 CTGGCCAATTTCCAAGACTATCTGTTTCGATGACAAGGCCAACGATTACGTCTACGAGTTC
GACCGGTTAAAGGTTCTGATAGACAAGCTACTGTTCCGGTTGCTAATGCAGATGCTCAAG
2761 TGGCGCGACAAGGTCCGCGCCCGCATCAAGGATCCGAAAGTTGCCGAGAAGCTCGCCCCC
ACCGCGCTGTTCCAGGCGCGGGCGTAGTTCCTAGGCTTTCAACGGCTCTTCGAGCGGGGG
2821 ATGAAGAAGCCGCATCCTTACGGAGCCAAGCGCCCTTCGCTGGAGCAGTGGTACTACGAG
TACTTCTTCGGCGTAGGAATGCCTCGGTTGCGGGGAAGCGACCTCGTCACCATGATGCTC
2881 ATCTTCAATCAGAACAACGTCACGCTGGTGGATGTCAACGAAACACCGGTGCTTCGCATC
TAGAAGTTAGTCTTGTTGCAGTGCGACCACCTACAGTTGCTTTGTGGCCACGAAGCGTAG
2941 ACCGAGAAAGGCATCGTGACCGCTGAGGGTGAAGCCGAATTCGACCTGATCGTGTTTCGCG

50

9/10

TGGCTCTTTCCGTAGCACTGGCGACTCCCCTTCGGCTTAAGCTGGACTAGCACAAAGCGC
3001 ACCGGCTTCGACGCAGTGACCGGGGGACTCACCAGCATCGACTTCGCAACAACCAGGGC
TGGCCGAAGCTGCGTCACTGGCCCCCTGAGTGGTCGTAGCTGAAGGCGTTGTTGGTCCCG
3061 CAGAGCTTCAAGGATGTGTGGTCTGACGGAATCCGCACCCAGCTGGGAGTGGCCACGGCA
GTCTCGAAGTTCCTACACACCAGACTGCCTTAGGCGTGGGTGACCCCTACCGGTGCCGT
3121 GGTTTTCCCAACTTGCTCTTTGGCTACGGACCTCAATCGCCTGCGGGCTTCTGCAACGGT
CCAAAAGGGTTGAACGAGAAACCGATGCCTGGAGTTAGCGGACGCCCCGAAGACGTTGCCA
3181 CCGTCGAGCGCCGAATACCAGGGCGATCTGCTGATCCAGCTGATGAACTACCTACGCGAC
GGCAGCTCGCGGCTTATGGTCCCGCTAGACGACTAGGTGACTACTTGATGGATGCGCTG
3241 AACACATCTCGCGCATCGAAGCCCAGTCCGAGGCACAGGAAGAATGGAGCAAGCTGATC
TTGTTGTAGAGCGCGTAGCTTCGGGTCAGGCTCCGTGTCCTTCTTACCTCGTTCGACTAG
3301 GCAGACTTCTGGGACAGCTCGCTGTTCCCCCGCGCAAAGTCCTGGTACCAAGGATCCAAC
CGTCTGAAGACCCTGTGAGCGACAAGGGGGCGCGTTTCAGGACCATGGTTCCTAGGTTG
3361 ATCCCGGGCAAGAAAGTCGAGAGCCTGAACTTCCCGCTGGGGCTGCCAACCTATATATCC
TAGGGCCCGTTCTTTCAGCTCTCGGACTTGAAGGGCGACCCCGACGGTTGGATATATAGG
3421 AAATTCAATGAAAGCGCTGAAAAAGGATATGCAGGCTTCTCGCTGGCCAGCTAAGACTCT
TTTAAGTTACTTTCGCGACTTTTTCTTATACGTCCGAAGAGCGACCGGTGATTCTGAGA
3481 GTTGTGCAACTCCTGGAGACAAGCATATGGGACGTGTAAATGACAAAGTGGTTCTCGTCA
CAACACGTTGAGGACCTCTGTTTCGTATACCCTGCACATTTACTGTTTCACCAAGAGCAGT
3541 CTGGCGGTGCCATGGGAATGGGCCTGACCCATTGCACCTTGCTGGCCCGCGAGGGGGCCA
GACCGCCACGGTACCCTTACCCGGACTGGGTAACGTGGAACGACCGGGCGCTCCCCCGGT
3601 CCGTTTACCTCAGCGACATGAATGAGGAACTGGGCCACCAAGCCGTAGCGGAGATCCGTC
GGCAAATGGAGTCGCTGTACTTACTCCTTGACCCGGTGGTTCGGCATCGCCTCTAGGCAG
3661 GGCAAGGTGGCAAGGCCCACTTCCTGCACCTGGACGTGACCAACGAGAATCACTGGACAG
CCGTTCCACCGTTCCGGGTGAAGGACGTGGACCTGCACTGGTTGCTCTTAGTGACCTGTC

Fi - 5E

10/10

3721 GTGCCGTGGACACCATCCTCGCCGAGTCCGACCGGCTGGACGCGCTGGTCAACAACGCCG
CACGGCACCTGTGGTAGGAGCGGCTCAGGCTGGCCGACCTGCGCGACCAGTTGTTGCGGC
3781 GCATTTTGACCCTCAAACCCGTTCAAGACACCAGCAACGAAGAATGGGATCGCATCTTTG
CGTAAAACTGGGAGTTTGGGCAAGTCCTGTGGTCGTTGCTTCTTACCCTAGCGTAGAAAC
3841 AAATCAATGTCCGCAGTGTGTTCTTGGGCACGCGAGCAGTGATCGAGCCCATGCGTAAAG
TTTAGTTACAGGCGTCACACAAGAACCCGTGCGCTCGTCACTAGCTCGGGTACGCATTTT
3901 CCCATAAAGGCTGCATCGTCAATGTGTCGTCCATCTACGGTCTGGTGGGTGCGCCGGGAG
GGGTATTTCCGACGTAGCAGTTACACAGCAGGTAGATGCCAGACCACCCACGCGGCCCTC
3961 CTGCGGCCTACGAGGCCTCCAAAGGTGCCGTGCGCTTGTTTACCAAGGCTTGCGCGGTGG
GACGCCGGATGCTCCGGAGGTTTCCACGGCACGCGAACAATGGTTCCGAACGCGCCACC
4021 ACCTGGCACCGTTCAATATTCGGGTCAACTCGGTCCACCCAGGGGTGATCGCAACACCCA
TGGACCGTGGCAAGTTATAAGCCCAGTTGAGCCAGGTGGGTCCCCACTAGCGTTGTGGGT
4081 TGACGCGAGCAGATCCTGGACGCCCCACAGAGCGCACGTGCGCTTCTGGGCCCCACGCTGC
ACTGCGTCGTCTAGGACCTGCGGGGTGTCTCGCGTGACGCGAAGACCCGGGGTGCGACG
4141 TGGGTCGTGCGGCCCAGCCGATGGAGGTCTCCAGGCGGTGCTTTTCCTGGTCTCGGACG
ACCCAGCACGCCGGGTCTGGCTACCTCCAGAGGGTCCGCCACGAAAAGGACCAGAGCCTGC
4201 AGGCCTCGTTCGTGCATGGCTCTGAACTCGTCGTCGACGGAGGGTATACGGCGAACTGAG
TCCGGAGCAAGCACGTACCGAGACTTGAGCAGCAGCTGCCTCCCATATGCCGCTTGACTC
4261 AAAGGAACATCCTCGGCATGC
TTTCCTTGTAGGAGCCGTACG

FE-5F

SEQUENCE LISTING

<110> NATIONAL RESEARCH COUNCIL OF CANADA
IWAKI, Hiroaki
HASEGAWA, Yoshie
LAU, Peter C.K.

<120> Cloning, Sequencing and Expression of a
Comamonas Cyclopentanone 1,2-Monooxygenase-Encoding Gene in
Escherichia coli

<130> 2139-16PCT

<150> US 60/218,842

<151> 2000-07-18

<160> 8

<170> FastSEQ for Windows Version 4.0

<210> 1

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<223> Primers for Comamonas sp.

<221> modified_base

<222> (3)...(3)

<223> I

<221> modified_base

<222> (6)...(6)

<223> I

<221> modified_base

<222> (12)...(12)

<223> I

<221> unsure

<222> (15)...(15)

<223> N = any base

<400> 1

acnacnatga cnacnatgac

20

<210> 2

<211> 17

<212> DNA

<213> Artificial Sequence

<220>

<223> Primers for Comamonas sp.

<221> modified_base

<222> (9)...(9)

<223> I

<400> 2

arrtgrtana rytgrta

17

<210> 3

<211> 31

<212> DNA

<213> Artificial Sequence

<220>

<223> Primers for Comamonas sp.

<400> 3

aaaaggcctg aacttcaatt atttaggaga c

31

<210> 4

<211> 32

<212> DNA

<213> Artificial Sequence

<220>

<223> Primers for Comamonas sp.

<400> 4

aaaactgcag gagttgcaca acagagtctt ag

32

<210> 5

<211> 550

<212> PRT

<213> Comamonas sp

<400> 5

Met	Thr	Thr	Met	Thr	Thr	Met	Thr	Thr	Glu	Gln	Leu	Gly	Met	Asn	Asn		
1				5					10					15			
Ser	Val	Asn	Asp	Lys	Leu	Asp	Val	Leu	Leu	Ile	Gly	Ala	Gly	Phe	Thr		
			20					25					30				
Gly	Leu	Tyr	Gln	Leu	Tyr	His	Leu	Arg	Lys	Leu	Gly	Tyr	Lys	Val	His		
		35					40				45						
Leu	Val	Asp	Ala	Gly	Ala	Asp	Ile	Gly	Gly	Ile	Trp	His	Trp	Asn	Cys		
	50					55				60							
Tyr	Pro	Gly	Ala	Arg	Val	Asp	Thr	His	Cys	Gln	Ile	Tyr	Gln	Tyr	Ser		
65					70					75					80		
Ile	Pro	Glu	Leu	Trp	Gln	Glu	Phe	Asn	Trp	Lys	Glu	Leu	Phe	Pro	Asn		
			85					90					95				
Trp	Ala	Gln	Met	Arg	Glu	Tyr	Phe	His	Phe	Ala	Asp	Lys	Lys	Leu	Asp		
			100					105					110				
Leu	Ser	Lys	Asp	Ile	Ser	Phe	Asn	Thr	Arg	Val	Gln	Ser	Ala	Val	Phe		
		115					120					125					
Asp	Glu	Gly	Thr	Arg	Glu	Trp	Thr	Val	Arg	Ser	Ile	Gly	His	Gln	Pro		
	130					135					140						
Ile	Gln	Ala	Arg	Phe	Val	Ile	Ala	Asn	Leu	Gly	Phe	Gly	Ala	Ser	Pro		
145					150					155					160		
Ser	Thr	Pro	Asn	Val	Asp	Gly	Ile	Glu	Thr	Phe	Lys	Gly	Gln	Trp	Tyr		
			165					170						175			
His	Thr	Ala	Leu	Trp	Pro	Gln	Glu	Gly	Val	Asn	Met	Ala	Gly	Lys	Arg		
			180					185					190				
Val	Ala	Ile	Ile	Gly	Thr	Gly	Ser	Ser	Gly	Val	Gln	Val	Ala	Gln	Glu		
		195				200						205					
Ala	Ala	Leu	Asp	Ala	Lys	Gln	Val	Thr	Val	Tyr	Gln	Arg	Thr	Pro	Asn		
	210					215						220					

```

Leu Ala Leu Pro Met His Gln Lys Gln Leu Ser Ala Glu Asp Asn Leu
225                230                235                240
Arg Met Lys Pro Glu Leu Pro Ala Ala Phe Glu Arg Arg Gly Lys Cys
                245                250                255
Phe Ala Gly Phe Asp Phe Asp Phe Ile Ala Lys Asn Ala Thr Glu Leu
                260                265                270
Ser Ala Ala Glu Arg Thr Glu Ile Leu Glu Glu Leu Trp Asn Ala Gly
                275                280                285
Gly Phe Arg Tyr Trp Leu Ala Asn Phe Gln Asp Tyr Leu Phe Asp Asp
                290                295                300
Lys Ala Asn Asp Tyr Val Tyr Glu Phe Trp Arg Asp Lys Val Arg Ala
305                310                315                320
Arg Ile Lys Asp Pro Lys Val Ala Glu Lys Leu Ala Pro Met Lys Lys
                325                330                335
Pro His Pro Tyr Gly Ala Lys Arg Pro Ser Leu Glu Gln Trp Tyr Tyr
                340                345                350
Glu Ile Phe Asn Gln Asn Asn Val Thr Leu Val Asp Val Asn Glu Thr
                355                360                365
Pro Val Leu Arg Ile Thr Glu Lys Gly Ile Val Thr Ala Glu Gly Glu
370                375                380
Ala Glu Phe Asp Leu Ile Val Phe Ala Thr Gly Phe Asp Ala Val Thr
385                390                395                400
Gly Gly Leu Thr Ser Ile Asp Phe Arg Asn Asn Gln Gly Gln Ser Phe
                405                410                415
Lys Asp Val Trp Ser Asp Gly Ile Arg Thr Gln Leu Gly Val Ala Thr
                420                425                430
Ala Gly Phe Pro Asn Leu Leu Phe Gly Tyr Gly Pro Gln Ser Pro Ala
                435                440                445
Gly Phe Cys Asn Gly Pro Ser Ser Ala Glu Tyr Gln Gly Asp Leu Leu
450                455                460
Ile Gln Leu Met Asn Tyr Leu Arg Asp Asn Asn Ile Ser Arg Ile Glu
465                470                475                480
Ala Gln Ser Glu Ala Gln Glu Glu Trp Ser Lys Leu Ile Ala Asp Phe
                485                490                495
Trp Asp Ser Ser Leu Phe Pro Arg Ala Lys Ser Trp Tyr Gln Gly Ser
                500                505                510
Asn Ile Pro Gly Lys Lys Val Glu Ser Leu Asn Phe Pro Leu Gly Leu
                515                520                525
Pro Thr Tyr Ile Ser Lys Phe Asn Glu Ser Ala Glu Lys Gly Tyr Ala
530                535                540
Gly Phe Ser Leu Ala Ser
545                550

```

<210> 6

<211> 549

<212> PRT

<213> Rhodococcus rhodochrous

<400> 6

```

Met Asn Gly Gln His Pro Arg Ser Val Val Thr Ala Pro Asp Ala Thr
1                5                10                15
Thr Gly Thr Thr Ser Tyr Asp Val Val Val Val Gly Ala Gly Ile Ala
                20                25                30
Gly Leu Tyr Ala Ile His Arg Phe Arg Ser Gln Gly Leu Thr Val Arg
                35                40                45
Ala Phe Glu Ala Ala Ser Gly Val Gly Gly Val Trp Tyr Trp Asn Arg
50                55                60
Tyr Pro Gly Ala Arg Cys Asp Val Glu Ser Ile Asp Tyr Ser Tyr Ser
65                70                75                80

```


Phe	Ser	Pro	Glu	Leu	Glu	Gln	Glu	Trp	Asn	Trp	Ser	Glu	Lys	Tyr	Ala	
				85					90					95		
Thr	Gln	Pro	Glu	Ile	Leu	Ala	Tyr	Leu	Glu	His	Val	Ala	Asp	Arg	Phe	
			100					105					110			
Asp	Leu	Arg	Arg	Asp	Ile	Arg	Phe	Asp	Thr	Arg	Val	Thr	Ser	Ala	Val	
		115					120					125				
Leu	Asp	Glu	Glu	Gly	Leu	Arg	Trp	Thr	Val	Arg	Thr	Asp	Arg	Gly	Asp	
	130					135					140					
Glu	Val	Ser	Ala	Arg	Phe	Leu	Val	Val	Ala	Ala	Gly	Pro	Leu	Ser	Asn	
145					150				155						160	
Ala	Asn	Thr	Pro	Ala	Phe	Asp	Gly	Leu	Asp	Arg	Phe	Thr	Gly	Asp	Ile	
				165				170						175		
Val	His	Thr	Ala	Arg	Trp	Pro	His	Asp	Gly	Val	Asp	Phe	Thr	Gly	Lys	
			180					185					190			
Arg	Val	Gly	Val	Ile	Gly	Thr	Gly	Ser	Ser	Gly	Ile	Gln	Ser	Ile	Pro	
		195					200					205				
Ile	Ile	Ala	Glu	Gln	Ala	Glu	Gln	Leu	Phe	Val	Phe	Gln	Arg	Ser	Ala	
	210					215					220					
Asn	Tyr	Ser	Ile	Pro	Ala	Gly	Asn	Val	Pro	Leu	Asp	Asp	Ala	Thr	Arg	
225					230				235						240	
Ala	Glu	Gln	Lys	Ala	Asn	Tyr	Ala	Glu	Arg	Arg	Arg	Leu	Ser	Arg	Glu	
			245					250						255		
Ser	Gly	Gly	Gly	Ser	Pro	His	Arg	Pro	His	Pro	Lys	Ser	Ala	Leu	Glu	
			260					265					270			
Val	Ser	Glu	Glu	Glu	Arg	Arg	Ala	Val	Tyr	Glu	Glu	Arg	Trp	Lys	Leu	
		275					280					285				
Gly	Gly	Val	Leu	Phe	Ser	Lys	Ala	Phe	Pro	Asp	Gln	Leu	Thr	Asp	Pro	
	290					295					300					
Ala	Ala	Asn	Asp	Thr	Ala	Arg	Ala	Phe	Trp	Glu	Glu	Lys	Ile	Arg	Ala	
305					310					315					320	
Val	Val	Asp	Asp	Pro	Ala	Val	Ala	Glu	Leu	Leu	Thr	Pro	Lys	Asp	His	
			325					330						335		
Ala	Ile	Gly	Ala	Lys	Arg	Ile	Val	Thr	Asp	Ser	Gly	Tyr	Tyr	Glu	Thr	
			340					345					350			
Tyr	Asn	Arg	Asp	Asn	Val	Glu	Leu	Val	Asp	Leu	Arg	Ser	Thr	Pro	Ile	
		355					360					365				
Val	Gly	Met	Asp	Glu	Thr	Gly	Ile	Val	Thr	Thr	Gly	Ala	His	Tyr	Asp	
	370					375					380					
Leu	Asp	Met	Ile	Val	Leu	Ala	Thr	Gly	Phe	Asp	Ala	Met	Thr	Gly	Ser	
385					390					395					400	
Leu	Asp	Lys	Leu	Glu	Ile	Val	Gly	Arg	Gly	Gly	Arg	Thr	Leu	Lys	Glu	
			405						410					415		
Thr	Trp	Ala	Ala	Gly	Pro	Arg	Thr	Tyr	Leu	Gly	Leu	Gly	Ile	Asp	Gly	
			420					425					430			
Phe	Pro	Asn	Phe	Phe	Asn	Leu	Thr	Gly	Pro	Gly	Ser	Pro	Ser	Val	Leu	
		435					440					445				
Ala	Asn	Met	Val	Leu	His	Ser	Glu	Leu	His	Val	Asp	Trp	Val	Ala	Asp	
	450					455					460					
Ala	Ile	Ala	Tyr	Leu	Asp	Ala	Arg	Gly	Ala	Ala	Gly	Ile	Glu	Gly	Thr	
465					470					475					480	
Pro	Glu	Ala	Val	Ala	Asp	Trp	Val	Glu	Glu	Cys	Arg	Asn	Arg	Ala	Glu	
			485					490						495		
Ala	Ser	Leu	Leu	Asn	Ser	Ala	Asn	Ser	Trp	Tyr	Leu	Gly	Ala	Asn	Ile	
			500					505					510			
Pro	Gly	Arg	Pro	Arg	Val	Phe	Met	Pro	Phe	Leu	Gly	Gly	Phe	Gly	Val	
		515					520					525				
Tyr	Arg	Glu	Ile	Ile	Thr	Glu	Val	Ala	Glu	Ser	Gly	Tyr	Lys	Gly	Phe	
	530					535					540					
Ala	Ile	Leu	Glu	Gly												
545																

<210> 7

<211> 543

<212> PRT

<213> Acinetobacter sp

<400> 7

Met	Ser	Gln	Lys	Met	Asp	Phe	Asp	Ala	Ile	Val	Ile	Gly	Gly	Gly	Phe
1				5					10					15	
Gly	Gly	Leu	Tyr	Ala	Val	Lys	Lys	Leu	Arg	Asp	Glu	Leu	Glu	Leu	Lys
			20					25					30		
Val	Gln	Ala	Phe	Asp	Lys	Ala	Thr	Asp	Val	Ala	Gly	Thr	Trp	Tyr	Trp
		35					40					45			
Asn	Arg	Tyr	Pro	Gly	Ala	Leu	Thr	Asp	Thr	Glu	Thr	His	Leu	Tyr	Cys
	50					55					60				
Tyr	Ser	Trp	Asp	Lys	Glu	Leu	Leu	Gln	Ser	Leu	Glu	Ile	Lys	Lys	Lys
65					70					75				80	
Tyr	Val	Gln	Gly	Pro	Asp	Val	Arg	Lys	Tyr	Leu	Gln	Gln	Val	Ala	Glu
				85					90					95	
Lys	His	Asp	Leu	Lys	Lys	Ser	Tyr	Gln	Phe	Asn	Thr	Ala	Val	Gln	Ser
			100					105					110		
Ala	His	Tyr	Asn	Glu	Ala	Asp	Ala	Leu	Trp	Glu	Val	Thr	Thr	Glu	Tyr
		115					120						125		
Gly	Asp	Lys	Tyr	Thr	Ala	Arg	Phe	Leu	Ile	Thr	Ala	Leu	Gly	Leu	Leu
	130					135					140				
Ser	Ala	Pro	Asn	Leu	Pro	Asn	Ile	Lys	Gly	Ile	Asn	Gln	Phe	Lys	Gly
145					150					155				160	
Glu	Leu	His	His	Thr	Ser	Arg	Trp	Pro	Asp	Asp	Val	Ser	Phe	Glu	Gly
				165					170					175	
Lys	Arg	Val	Gly	Val	Ile	Gly	Thr	Gly	Ser	Thr	Gly	Val	Gln	Val	Ile
			180					185					190		
Thr	Ala	Val	Ala	Pro	Leu	Ala	Lys	His	Leu	Thr	Val	Phe	Gln	Arg	Ser
		195					200					205			
Ala	Gln	Tyr	Ser	Val	Pro	Ile	Gly	Asn	Asp	Pro	Leu	Ser	Glu	Glu	Asp
	210					215					220				
Val	Lys	Lys	Ile	Lys	Asp	Asn	Tyr	Asp	Lys	Ser	Leu	Gly	Trp	Cys	Met
225					230					235				240	
Asn	Ser	Ala	Leu	Ala	Phe	Ala	Leu	Asn	Glu	Ser	Thr	Val	Pro	Ala	Met
				245					250					255	
Ser	Val	Ser	Ala	Glu	Glu	Arg	Lys	Ala	Val	Phe	Glu	Lys	Ala	Trp	Gln
			260				265						270		
Thr	Gly	Gly	Gly	Phe	Arg	Phe	Met	Phe	Glu	Thr	Phe	Gly	Asp	Ile	Ala
		275					280					285			
Thr	Asn	Met	Glu	Ala	Asn	Ile	Glu	Ala	Gln	Asn	Phe	Ile	Lys	Gly	Lys
	290					295					300				
Ile	Ala	Glu	Ile	Val	Lys	Asp	Pro	Ala	Ile	Ala	Gln	Lys	Leu	Met	Pro
305					310					315				320	
Gln	Asp	Leu	Tyr	Ala	Lys	Arg	Pro	Leu	Cys	Asp	Ser	Gly	Tyr	Tyr	Asn
				325					330					335	
Thr	Phe	Asn	Arg	Asp	Asn	Val	Arg	Leu	Glu	Asp	Val	Lys	Ala	Asn	Pro
			340					345					350		
Ile	Val	Glu	Ile	Thr	Glu	Asn	Gly	Val	Lys	Leu	Glu	Asn	Gly	Asp	Phe
		355					360					365			
Val	Glu	Leu	Asp	Met	Leu	Ile	Cys	Ala	Thr	Gly	Phe	Asp	Ala	Val	Asp
	370					375					380				
Gly	Asn	Tyr	Val	Arg	Met	Asp	Ile	Gln	Gly	Lys	Asn	Gly	Leu	Ala	Met
385					390					395				400	
Lys	Asp	Tyr	Trp	Lys	Glu	Gly	Pro	Ser	Ser	Tyr	Met	Gly	Val	Thr	Val
				405					410					415	

```

Asn Asn Tyr Pro Asn Met Phe Met Val Leu Gly Pro Asn Gly Pro Phe
      420                      425                      430
Thr Asn Leu Pro Pro Ser Ile Glu Ser Gln Val Glu Trp Ile Ser Asp
      435                      440                      445
Thr Ile Gln Tyr Thr Val Glu Asn Asn Val Glu Ser Ile Glu Ala Thr
      450                      455                      460
Lys Glu Ala Glu Glu Gln Trp Thr Gln Thr Cys Ala Asn Ile Ala Glu
465                      470                      475                      480
Met Thr Leu Phe Pro Lys Ala Gln Ser Trp Ile Phe Gly Ala Asn Ile
      485                      490                      495
Pro Gly Lys Lys Asn Thr Val Tyr Phe Tyr Leu Gly Gly Leu Lys Glu
      500                      505                      510
Tyr Arg Thr Cys Ala Ser Asn Cys Lys Asn His Ala Tyr Glu Gly Phe
      515                      520                      525
Asp Ile Gln Leu Gln Arg Ser Asp Ile Lys Gln Pro Ala Asn Ala
      530                      535                      540

```

<210> 8

<211> 4281

<212> DNA

<213> Comamonas sp

<220>

<221> CDS

<222> (1822)...(3471)

<400> 8

```

gcatgcgata gcgggcacgtt ggtggggcggg cgagcgcggc caccgagcgcg tctctcgcctc 60
caccgacatc acatcgccgg tcaccggcga gaacattggc aagatcctgc gcttttcgcc 120
gctgaagccg cagggctacg agagaatttt tcagtccggc ctggcggact cgaccagca 180
cacccccagg atggtgctgt tcggacagtt cctgtcctgt gatccagaga ccatcaacac 240
cctggagaca ttggagcggg tcgcgcgcgc cgacgtcaat gtcttgctgc atggagagac 300
gggcacgggg aaagagctga tagctcgcca cattcatgtc gcgagccgcc ggcgggatgc 360
gccctacctc gccatcaact gcgggggcaat cagctcggag ttgctggaga gtactttttt 420
tggttatgtg cgcggagcat tctccggggc agatcccaag ggccgcgccg gctactttga 480
atcgggtggg gaaggcacct tgtttctgga cgagatcggc gagctgccct tggccatgca 540
ggcggcattg ttgcgcgtgc tggaagacgg aagctttctg cgtgtgggct catcgacccc 600
gcagcgcgcc tcgtgccgca ttatcgcggc caccgaccgc aatctggagg aactcatcgc 660
acagggcctg tttcgccagg atctctacta ccgtctcaag atcgttcaga aaaggctcaa 720
gccgatacgg gaacgcacct gtgacatcgc cttgctggca gagcaattca aatctgcact 780
ggcgcaaaag caccagatcc cgaacgtgca gatccacccc gaggccatgg cagtcattga 840
gcgctaccag tggccggggc atgcgcgcga aatccgcaat gtgatggaag ccgccctgat 900
ctgctccgat ggtgaaatca cgctcgccag cctgcccccg gaagtatctg agaactccac 960
ctatcctctg caaagccggg tggccgaagc gaatagcgag atgcctcccg tctccggcaa 1020
cgactatgaa cggcaactca tcgtgggttt gctgcgcaag taccgaaagg tcaatcatgt 1080
ggcgagggcc ctgggcctcg cgcgctccac gctctatcgg aaattcgag acttggggcat 1140
tgaccaacgc gagtatgtca atgatctgtc cgattaacac tgctgcgcc tgcgcttgcg 1200
gcaacgcagt atcgtcgga cgcgggcaac gctaaaggca gccgccgtcg gcatcggttg 1260
ccgcccgggt ttgtttgtcg gctgagaccg cctgcagcac cagccctcc attcctcgtg 1320
tctagatctc cttgcaacgc tgtcacagcg tgcttttttc gtggcgcggg acttctcgcg 1380
atgagggcct gcaagggcgg cgcatacaaa caagcgccct gtcctcagga cgtttgcgcc 1440
gtcgctggct atgggcagtc tgcattgatc gcttgggctt gcccaccatg cgcgcgactg 1500
cgctctttcc tgttggggcg atcgtcatcc cgaccttcc gaaatccatt tccacagggc 1560
cggcctgagc cagacagcga tcggcaggct ctgcacatct ggacagtggg gcgctacacc 1620
acaggacaca agtgtccggc gcgtggcacg cagcggcctt tgctgcgcac gatgcgtgct 1680
tttggcacat ggaaaccctc ggtttttccg aaccagagac ctggcacaag tcttgataac 1740
tcctgagtga ctgatgcaac gcatctcagt ccgggagcgt ggacgacacg cccctgaact 1800
tcaattattt aggagacca t atg acc acc atg acc acc atg acc acc gaa 1851

```

Met Thr Thr Met Thr Thr Met Thr Thr Glu

1

5

10

caa ctc ggc atg aac aac tct gtc aat gac aag ctt gac gtt ttg ctc	1899
Gln Leu Gly Met Asn Asn Ser Val Asn Asp Lys Leu Asp Val Leu Leu	
15 20 25	
atc ggc gcc ggc ttc acc ggt ctc tac cag ctc tat cac ctg cgc aag	1947
Ile Gly Ala Gly Phe Thr Gly Leu Tyr Gln Leu Tyr His Leu Arg Lys	
30 35 40	
ctg ggc tac aag gtt cat ctc gtc gac gcc ggt gcc gat att ggc ggg	1995
Leu Gly Tyr Lys Val His Leu Val Asp Ala Gly Ala Asp Ile Gly Gly	
45 50 55	
atc tgg cat tgg aac tgc tac ccc gga gcg cgt gtg gat acc cac tgc	2043
Ile Trp His Trp Asn Cys Tyr Pro Gly Ala Arg Val Asp Thr His Cys	
60 65 70	
cag atc tac cag tac tcc att cca gag ttg tgg cag gag ttc aac tgg	2091
Gln Ile Tyr Gln Tyr Ser Ile Pro Glu Leu Trp Gln Glu Phe Asn Trp	
75 80 85 90	
aaa gag ctg ttc cct aac tgg gcg caa atg cgc gag tat ttc cat ttt	2139
Lys Glu Leu Phe Pro Asn Trp Ala Gln Met Arg Glu Tyr Phe His Phe	
95 100 105	
gcc gac aag aag ctc gac ctg agc aag gac atc agc ttc aac acc cgt	2187
Ala Asp Lys Lys Leu Asp Leu Ser Lys Asp Ile Ser Phe Asn Thr Arg	
110 115 120	
gtg cag tcg gcc gtc ttt gac gaa ggc aca cgc gaa tgg acg gta cgc	2235
Val Gln Ser Ala Val Phe Asp Glu Gly Thr Arg Glu Trp Thr Val Arg	
125 130 135	
tcg atc gga cac cag ccg atc cag gcc agg ttc gtc atc gcc aac ctt	2283
Ser Ile Gly His Gln Pro Ile Gln Ala Arg Phe Val Ile Ala Asn Leu	
140 145 150	
ggc ttc ggt gcc agc ccc agc acg ccc aat gtc gat ggc atc gag aca	2331
Gly Phe Gly Ala Ser Pro Ser Thr Pro Asn Val Asp Gly Ile Glu Thr	
155 160 165 170	
ttc aaa ggc cag tgg tat cac act gct ctg tgg ccc cag gaa ggc gtg	2379
Phe Lys Gly Gln Trp Tyr His Thr Ala Leu Trp Pro Gln Glu Gly Val	
175 180 185	
aac atg gcc ggc aag cgc gtg gcc atc att ggc acc ggc tcc agc ggg	2427
Asn Met Ala Gly Lys Arg Val Ala Ile Ile Gly Thr Gly Ser Ser Gly	
190 195 200	
gtc cag gtc gcc cag gag gct gcc ctt gat gcg aaa cag gtg acg gtg	2475
Val Gln Val Ala Gln Glu Ala Ala Leu Asp Ala Lys Gln Val Thr Val	
205 210 215	
tac cag cgc acc ccc aac ctg gcc ttg ccc atg cat cag aag cag ctc	2523
Tyr Gln Arg Thr Pro Asn Leu Ala Leu Pro Met His Gln Lys Gln Leu	
220 225 230	
agc gcc gag gac aat ctg cgc atg aag ccc gag ctt ccc gca gcg ttc	2571
Ser Ala Glu Asp Asn Leu Arg Met Lys Pro Glu Leu Pro Ala Ala Phe	
235 240 245 250	

gag aga cgc ggc aag tgc ttc gcc ggc ttc gac ttc gac ttc atc gcc	2619
Glu Arg Arg Gly Lys Cys Phe Ala Gly Phe Asp Phe Asp Phe Ile Ala	
255 260 265	
aag aac gcg acc gag ctg tcc gct gcg gag cgc aca gag atc ttg gaa	2667
Lys Asn Ala Thr Glu Leu Ser Ala Ala Glu Arg Thr Glu Ile Leu Glu	
270 275 280	
gag ctg tgg aac gcc ggc ggc ttc cgc tac tgg ctg gcc aat ttc caa	2715
Glu Leu Trp Asn Ala Gly Gly Phe Arg Tyr Trp Leu Ala Asn Phe Gln	
285 290 295	
gac tat ctg ttc gat gac aag gcc aac gat tac gtc tac gag ttc tgg	2763
Asp Tyr Leu Phe Asp Asp Lys Ala Asn Asp Tyr Val Tyr Glu Phe Trp	
300 305 310	
cgc gac aag gtc cgc gcc cgc atc aag gat ccg aaa gtt gcc gag aag	2811
Arg Asp Lys Val Arg Ala Arg Ile Lys Asp Pro Lys Val Ala Glu Lys	
315 320 325 330	
ctc gcc ccc atg aag aag ccg cat cct tac gga gcc aag cgc cct tcg	2859
Leu Ala Pro Met Lys Lys Pro His Pro Tyr Gly Ala Lys Arg Pro Ser	
335 340 345	
ctg gag cag tgg tac tac gag atc ttc aat cag aac aac gtc acg ctg	2907
Leu Glu Gln Trp Tyr Tyr Glu Ile Phe Asn Gln Asn Asn Val Thr Leu	
350 355 360	
gtg gat gtc aac gaa aca ccg gtg ctt cgc atc acc gag aaa ggc atc	2955
Val Asp Val Asn Glu Thr Pro Val Leu Arg Ile Thr Glu Lys Gly Ile	
365 370 375	
gtg acc gct gag ggt gaa gcc gaa ttc gac ctg atc gtg ttc gcg acc	3003
Val Thr Ala Glu Gly Glu Ala Glu Phe Asp Leu Ile Val Phe Ala Thr	
380 385 390	
ggc ttc gac gca gtg acc ggg gga ctc acc agc atc gac ttc cgc aac	3051
Gly Phe Asp Ala Val Thr Gly Gly Leu Thr Ser Ile Asp Phe Arg Asn	
395 400 405 410	
aac cag ggc cag agc ttc aag gat gtg tgg tct gac gga atc cgc acc	3099
Asn Gln Gly Gln Ser Phe Lys Asp Val Trp Ser Asp Gly Ile Arg Thr	
415 420 425	
cag ctg gga gtg gcc acg gca ggt ttt ccc aac ttg ctc ttt ggc tac	3147
Gln Leu Gly Val Ala Thr Ala Gly Phe Pro Asn Leu Leu Phe Gly Tyr	
430 435 440	
gga cct caa tcg cct gcg ggc ttc tgc aac ggt ccg tcg agc gcc gaa	3195
Gly Pro Gln Ser Pro Ala Gly Phe Cys Asn Gly Pro Ser Ser Ala Glu	
445 450 455	
tac cag ggc gat ctg ctg atc cag ctg atg aac tac cta cgc gac aac	3243
Tyr Gln Gly Asp Leu Leu Ile Gln Leu Met Asn Tyr Leu Arg Asp Asn	
460 465 470	
aac atc tcg cgc atc gaa gcc cag tcc gag gca cag gaa gaa tgg agc	3291
Asn Ile Ser Arg Ile Glu Ala Gln Ser Glu Ala Gln Glu Glu Trp Ser	
475 480 485 490	

aag ctg atc gca gac ttc tgg gac agc tcg ctg ttc ccc cgc gca aag	3339
Lys Leu Ile Ala Asp Phe Trp Asp Ser Ser Leu Phe Pro Arg Ala Lys	
495 500 505	
tcc tgg tac caa gga tcc aac atc ccg ggc aag aaa gtc gag agc ctg	3387
Ser Trp Tyr Gln Gly Ser Asn Ile Pro Gly Lys Lys Val Glu Ser Leu	
510 515 520	
aac ttc ccg ctg ggg ctg cca acc tat ata tcc aaa ttc aat gaa agc	3435
Asn Phe Pro Leu Gly Leu Pro Thr Tyr Ile Ser Lys Phe Asn Glu Ser	
525 530 535	
gct gaa aaa gga tat gca ggc ttc tcg ctg gcc agc taagactctg	3481
Ala Glu Lys Gly Tyr Ala Gly Phe Ser Leu Ala Ser	
540 545 550	
ttgtgcaact cctggagaca agcatatggg acgtgtaaat gacaaagtgg ttctcgtcac	3541
tggcgggtgcc atgggaatgg gcctgaccca ttgcaccttg ctggcccgcg agggggccac	3601
cgtttacctc agcgacatga atgaggaact gggccaccaa gccgtagcgg agatccgtcg	3661
gcaaggtggc aaggcccact tcctgcacct ggacgtgacc aacgagaatc actggacagg	3721
tgccgtggac accatcctcg ccgagtccga ccggctggac gcgctggtca acaacgccgg	3781
cattttgacc ctcaaaccg ttcaggacac cagcaacgaa gaatgggatc gcatctttga	3841
aatcaatgtc cgcagtgtgt tcttggggcac gcgagcagtg atcgagccca tgcgtaaagc	3901
ccataaaggc tgcatacgtca atgtgtcgtc catctacggt ctggtgggtg cgccgggagc	3961
tgcggcctac gaggcctcca aagggtgccgt gcgcttggtt accaaggctt gcgcgggtgga	4021
cctggcaccg ttcaatatc ggggtcaactc ggtccaccca ggggtgatcg caacacccat	4081
gacgcagcag atcctggacg cccacacagag cgcacgtgcg cttctggggc ccacgctgct	4141
gggtcgtgcg gccagccga tggaggtctc ccaggcgggtg cttttcctgg tctcggacga	4201
ggcctcgttc gtgcatggct ctgaactcgt cgtcgacgga gggatatcgg cgaactgaga	4261
aaggaacatc ctcggcatgc	4281

**This Page is Inserted by IFW Indexing and Scanning
Operations and is not part of the Official Record**

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- ☐ **BLACK BORDERS**
- ☐ **IMAGE CUT OFF AT TOP, BOTTOM OR SIDES**
- ☒ **FADED TEXT OR DRAWING**
- ☐ **BLURRED OR ILLEGIBLE TEXT OR DRAWING**
- ☐ **SKEWED/SLANTED IMAGES**
- ☐ **COLOR OR BLACK AND WHITE PHOTOGRAPHS**
- ☐ **GRAY SCALE DOCUMENTS**
- ☒ **LINES OR MARKS ON ORIGINAL DOCUMENT**
- ☐ **REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY**
- ☐ **OTHER:** _____

IMAGES ARE BEST AVAILABLE COPY.

As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.